

Inverse Method to Estimate Microbial Inactivation Kinetic Parameters in Conduction-Heated Canned Foods

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Microbial Inactivation Kinetic Parameters

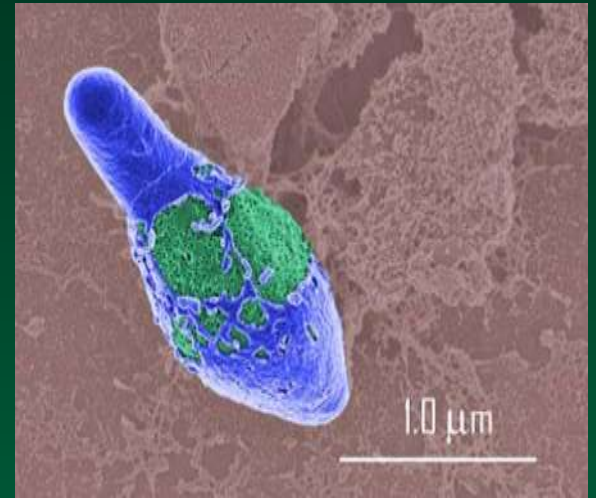
Needed to determine the time and temperature treatment for canned foods.

- ❑ Food containers used annually:
 - 22 billion steel cans
 - 75 billion glass jars
- ❑ Low-acid canned foods need $\sim 121^{\circ}\text{C}$ for 20-60 min. to inactivate *Clostridium botulinum*.



Clostridium botulinum

- ❑ Most potent natural toxin known
- ❑ One-millionth of a gram of toxin can kill an adult
- ❑ Spores cannot grow and cannot produce toxin when $\text{pH} \leq 4.6$
- ❑ Canning regulations are based on *C. bot*



How microbial kinetic parameters are usually estimated

- For high-moisture foods at constant temperature and constant moisture
 - Typical methods: capillary tubes for liquids
 - Assumes minimal lag time, no temp. gradients
 - Easy set-up and straightforward math

Example for high-moisture foods

- First-order model with Arrhenius rate constant. Temp. constant.

$$\log_{10}(N / N_0) = -t / D_r$$

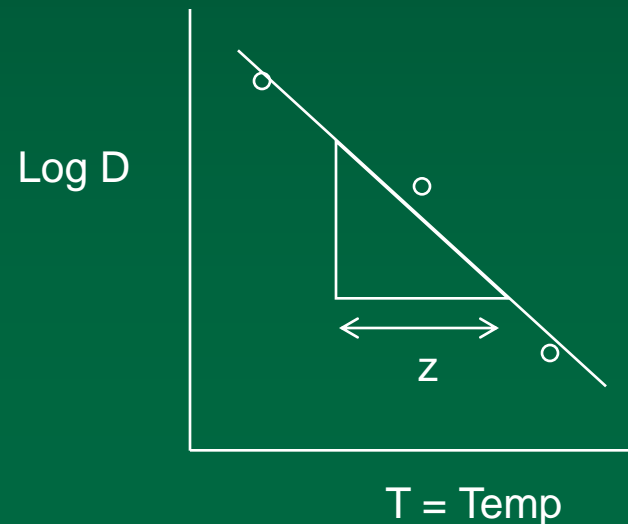
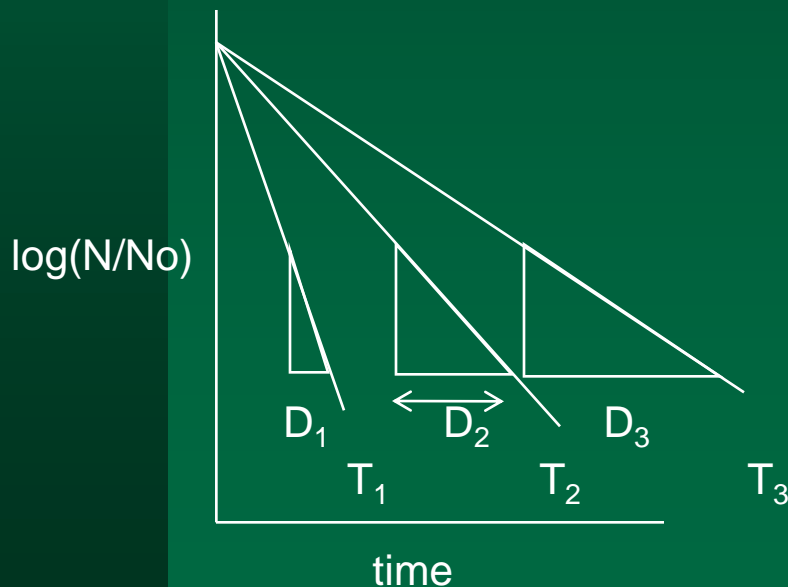
$$D = D_r 10^{\frac{-(T-T_r)}{z}}$$

N: # of m.o.'s at time t

N₀: initial number m.o.'s

D_r = inverse rate, min

z = temp change to cause
10-fold increase in D



Disadvantages of isothermal methods

- ❑ Limited temperature range gives lower statistical confidence
- ❑ More experiments required
- ❑ Unavoidable thermal lags
- ❑ Small samples may not be large enough for high concentration of m.o.'s

Advantages of dynamic nonisothermal expts

- ❑ Can estimate parameters with one experiment
- ❑ Covers entire temp range
- ❑ No thermal lag, can handle solid foods
- ❑ Sample can be any size, as long as temperature gradient is known
- ❑ Represents most true food processes
- ❑ Disadvantage: Math is more complex

For Conduction-heated foods

- ❑ First-order model with Arrhenius rate constant. Temp. and moisture changing with time.

- ❑ First-order $\frac{dN}{dt} = -N / D$

- ❑ Reciprocal Rate: $D = D_r 10^{\frac{-(T-T_r)}{z}}$

- ❑ Integrate:

$$\log(N / N_0) = -\left(\frac{1}{D}\right) \int_0^t 10^{\left(\frac{T(t)-T_r}{z}\right)} dt$$

- ❑ This is the microbial survival ratio at any one point in the can.
- ❑ Nonlinear regression with Excel Solver or Matlab to estimate D_r, z

Objectives

To develop a method to:

1. Estimate *kinetic parameters* for microbial inactivation in conduction-heated foods;
2. Compute confidence contours for the two kinetic parameters, D_r and z

Materials and Methods

Overview of the process

Inoculation of
canned pea puree
with *B. Stearothermophilus* spores

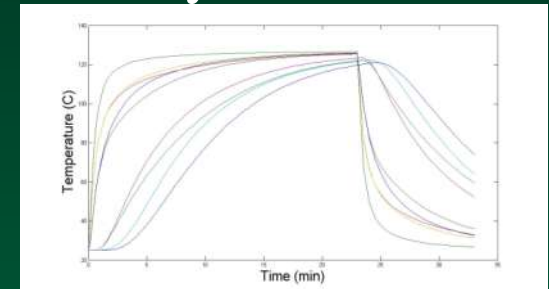


$N_0 = 9 \times 10^5$ CFU/mL
(Welt, et al.
1997 J. Food
Sci)

Retorting at
different times
at 104, 112, 120°C



Temperatures inside
can predicted by
analytical solns



Kinetic parameter
estimation



Plating of spores
to determine N



Materials

- Can size = 6.02 cm dia x 3.48 cm height (squat “tuna” can)
- Finite cylinder with boundary conditions of the 3rd kind: $h \approx 5500$ W/m²°C (steam)
- Uniform initial temp = 0°C
- $k = 0.68$ W/m°C, $C_p = 4100$ J/g°C

Analytical Solution to heating finite cylinder

- Analytic solution is product of infinite slab and cylinder for heating

$$\frac{T(r, \xi, t) - T_s}{T_i - T_s} = \sum_{m=1}^{\infty} \frac{2 \left(\frac{hl}{k} \right) \cos(\lambda_m \xi) \sec(\lambda_m) \exp \left[-\lambda_m^2 \left(\frac{\alpha t}{l^2} \right) \right]}{\left(\frac{hl}{k} \right) \left(\frac{hl}{k} + 1 \right) + \lambda_m^2} \quad \left. \vphantom{\sum_{m=1}^{\infty}} \right\} \text{Slab}$$

$$\times \sum_{n=1}^{\infty} \frac{2 \left(\frac{hR}{k} \right) J_0(\lambda_n r) \exp \left[-\lambda_n^2 \left(\frac{\alpha t}{R^2} \right) \right]}{\left[\left(\frac{hR}{k} \right)^2 + \lambda_n^2 \right] J_0(\lambda_n)} \quad \left. \vphantom{\sum_{n=1}^{\infty}} \right\} \text{Cylinder}$$

Analytical Solution to cooling finite cylinder

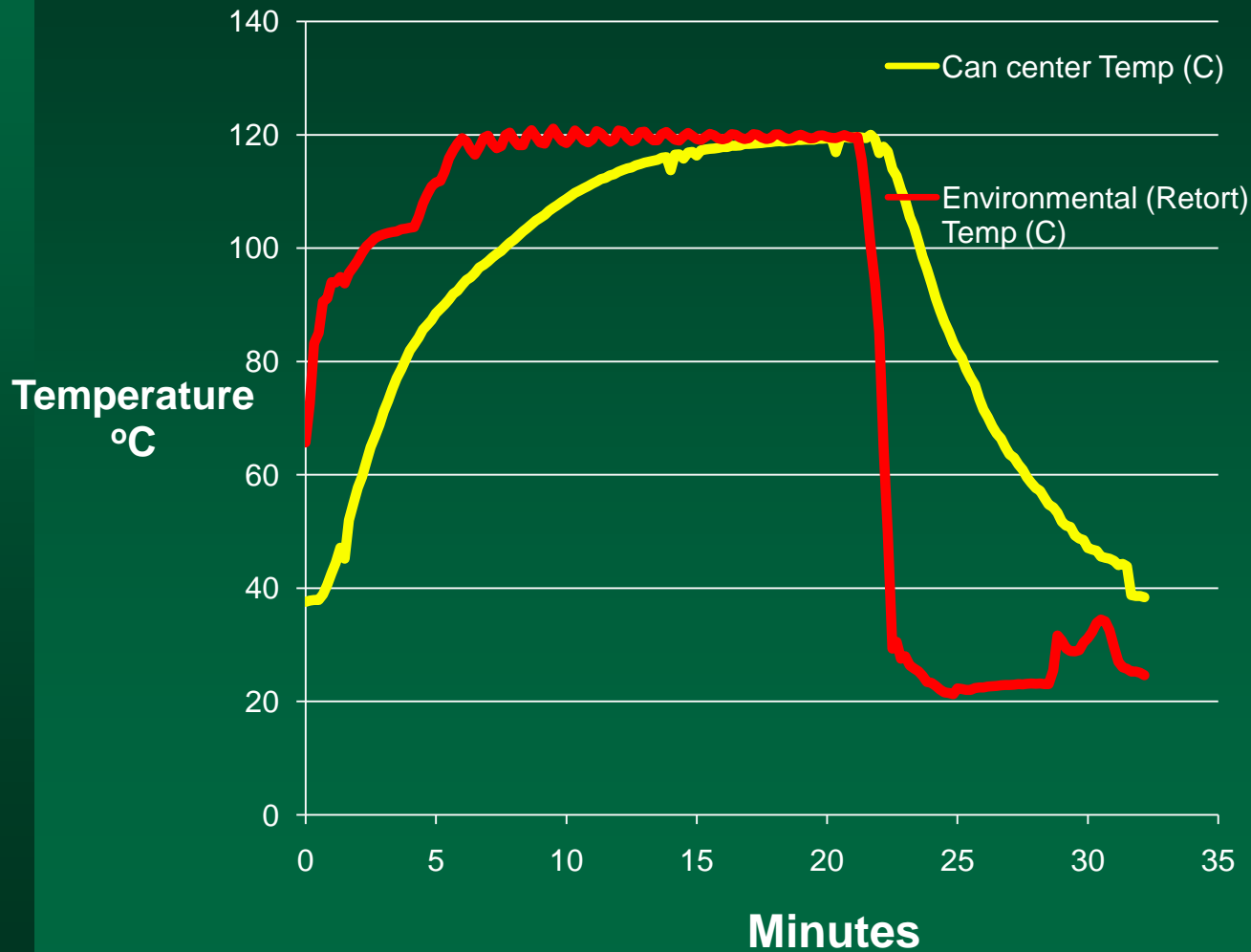
- Analytic solution when heating stops (non-uniform temp initial condition) and cooling begins . Assumes $h = \infty$ at boundary

$$\frac{T(r, \xi, \tau) - T_w}{T_s - T_i} = 4 \sum_{j=0}^{\infty} \sum_{i=0}^{\infty} \frac{(-1)^i \exp\left[-\left(\gamma^2 \left(i + \frac{1}{2}\right)^2 \pi^2 + \omega_j^2\right) \tau_w\right] J_0(\omega_j r)}{\left(i + \frac{1}{2}\right) \pi \omega_j J_1(\omega_j)}$$

$$\times \cos\left[\left(i + \frac{1}{2}\right) \pi \xi\right] \left\{ \left(\frac{T_s - T_w}{T_s - T_i}\right) - \exp\left[-\left(\omega_j^2 + \left(i + \frac{1}{2}\right)^2 \pi^2 \gamma^2\right) \tau\right] \right\}$$

Lenz, M. K., & Lund, D. B. (1977a). The lethality-Fourier number method: experimental verification of a model for calculating temperature profiles and lethality in conduction-heating canned foods. *Journal of Food Science*, 42(4), 989–996, 1001.

Typical measured temperature during steam retort processing



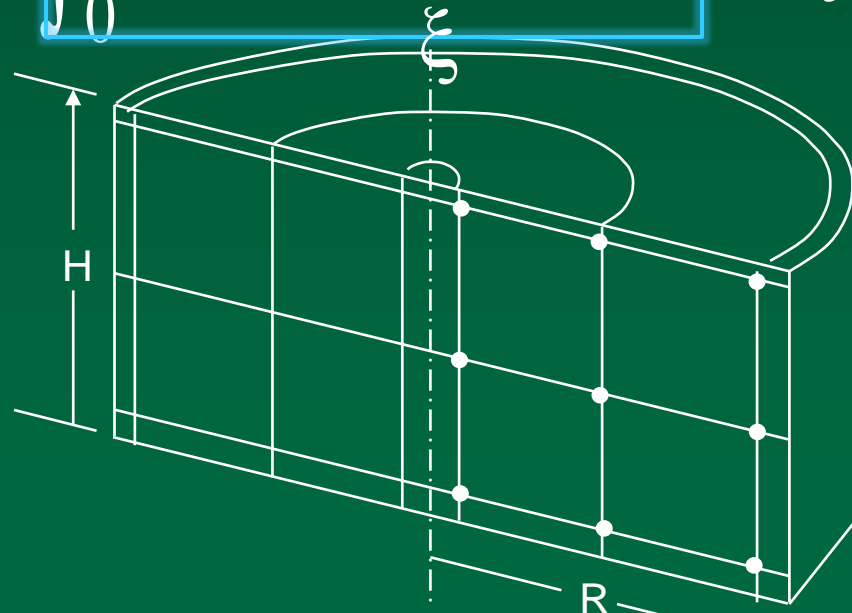
Model Formulation

Inverse Problem: Kinetic Parameters of Microbial inactivation

Calculated mass-average retention =

$$\left(\bar{N} / N_0\right)_{\text{model}} = 2 \int_0^H \int_0^R 10^{-\left(\frac{1}{D_r}\right) \beta(r, \xi, t)} r dr d\xi$$

- 3-point Gauss integration



Methods: Different Heating Treatments

- Heated canned pea puree in retort at three constant retort temperatures (T_{∞}): 104, 112, 120°C
- Heating times: 0-305 min
- Standard microbial techniques to measure mass-average microbial retention in each can =

$$(\bar{N} / N_0)_{observed}$$

Estimation of Kinetic Parameters

- Minimize Sum of Squares of errors

$$SSQ = \sum_{i=1}^n \left[\log \left(\frac{\bar{N}}{N_0} \right)_{\text{obs}, i} - \log \left(\frac{\bar{N}}{N_0} \right)_{\text{model}, i} \right]^2$$

Scaled Sensitivity Coefficients

□ Model: $\log(N / N_0) = -\left(\frac{1}{D_r}\right) \int_0^t 10^{\left(\frac{T(t)-T_r}{z}\right)} dt = -\left(\frac{1}{D_r}\right) \beta$

$$D_r \frac{\partial \log(N / N_0)}{\partial D_r} = D_r \left(\frac{\beta}{D_r^2} \right) = -\log(N / N_0)$$

$$\begin{aligned} z \frac{\partial \log(N / N_0)}{z} &= z \left(\frac{1}{D_r z^2} \right) \int_0^t (T(r, z, t) - T_r) 10^{\left(\frac{T(t)-T_r}{z}\right)} dt \\ &= \frac{\beta'}{D_r z} \end{aligned}$$

Parameter Joint Confidence Region

□ Motulsky iteration method

$$SS_{all-fixed} = SS_{best-fit} \left(\frac{p}{n-p} F(p, n-p) + 1 \right)$$

- n: number of data
- p: number of parameters
- F: statistical F distribution
- Orders of magnitude more computationally intensive than elliptical approximation

Asymptotic Confidence Bands

- Supplied by nlinfit in Matlab®

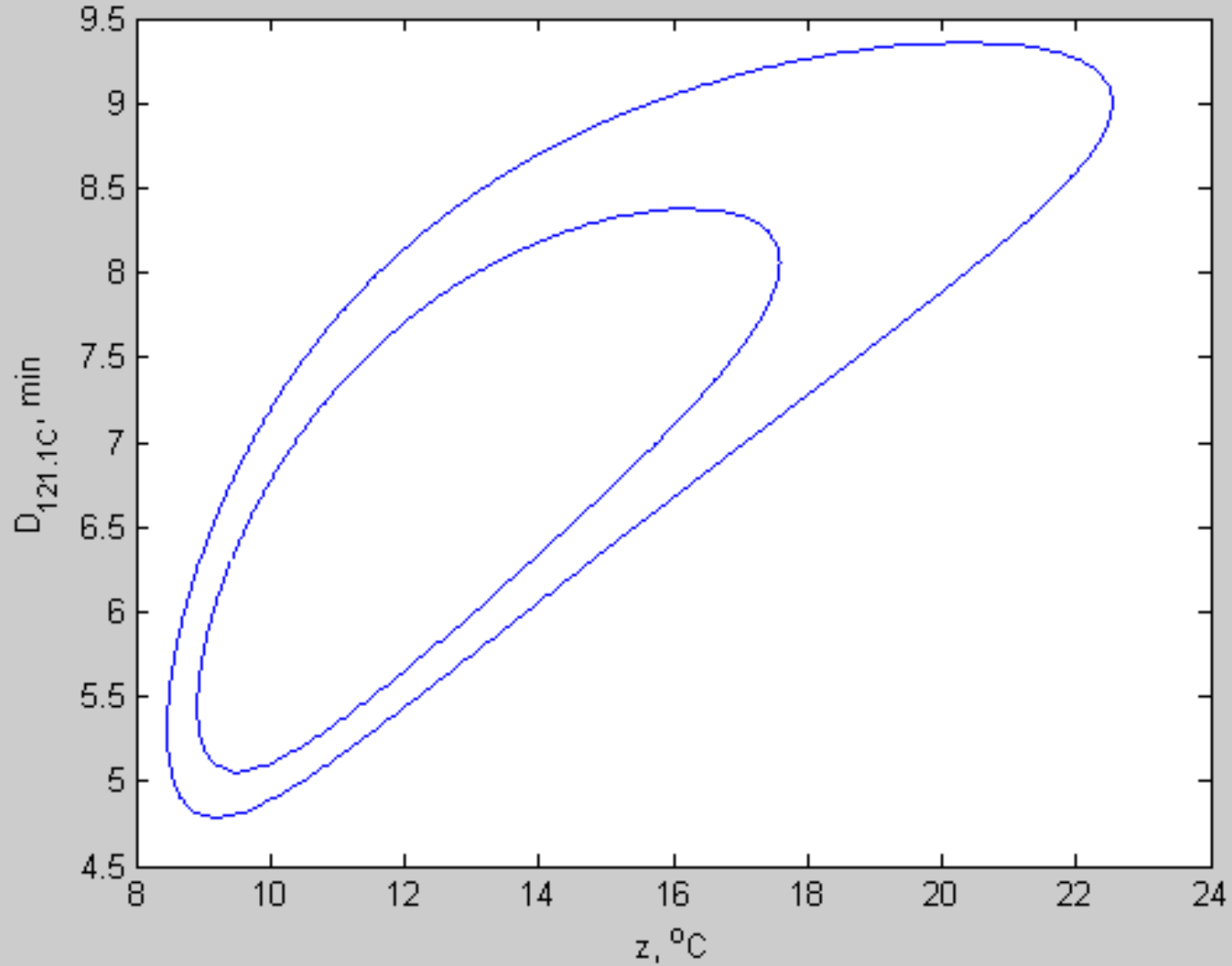
Results

Results for 2 parameters estimation

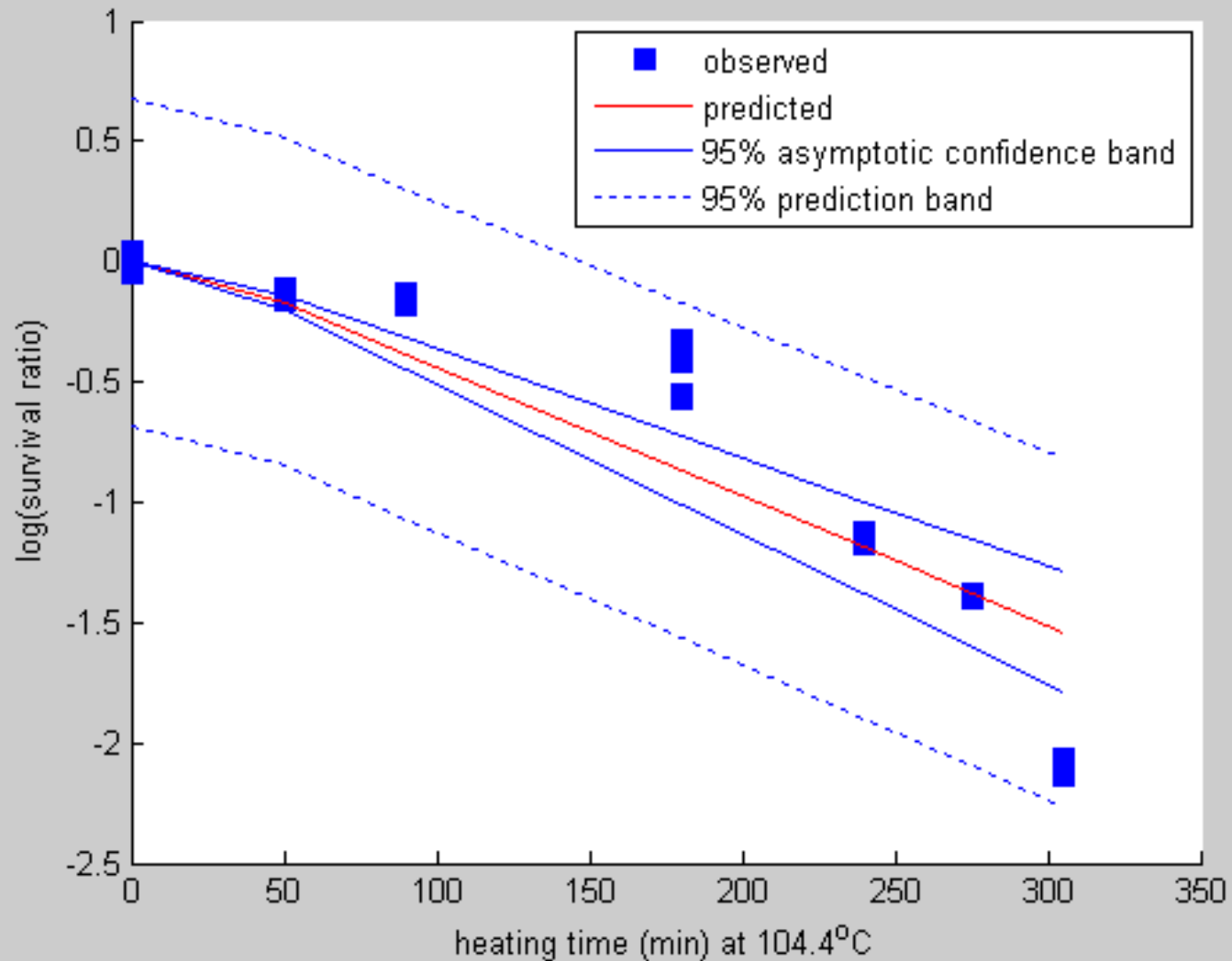
$$\left(\frac{\bar{N}}{N_o} \right)_{\text{pred}} = 2 \iint 10^{\frac{1}{D_r} \int_0^t 10^{\left(\frac{T(r,z,t) - T_r}{z} \right) dt} r dr d\xi$$

root mean square error	Number of data	parameter estimate	standard error	correlation coefficient $\rho_{D_r, z}$	95% asymptotic confidence interval
0.225	24	$D_{121.1^\circ\text{C}} = 6.67 \text{ min}$ $z = 11.74^\circ\text{C}$	0.421 min^{-1} 0.900 $^\circ\text{C}$	0.529 $(T_r = 381 \text{ K})$	(5.80, 7.54) (9.88, 13.61)

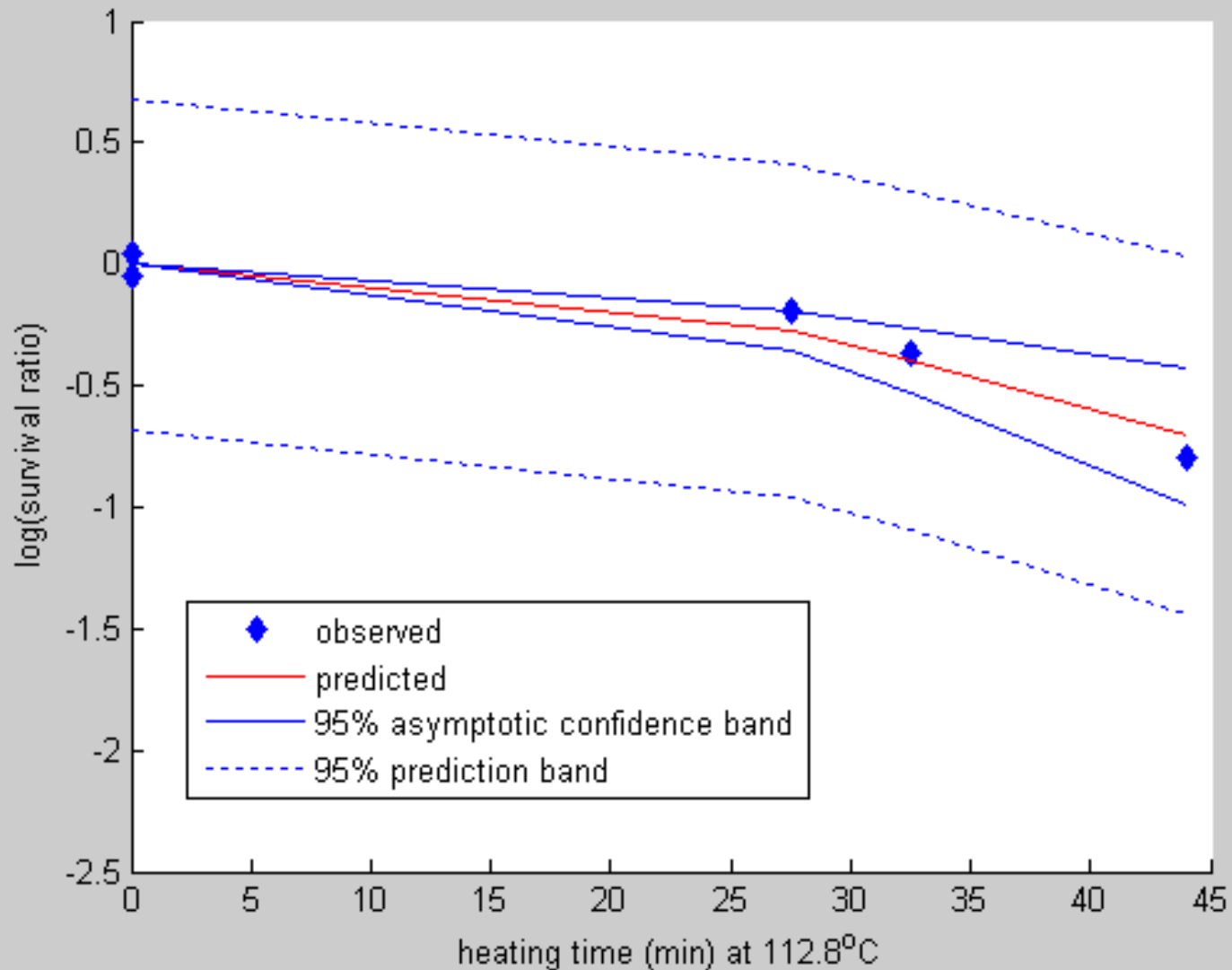
95%(inner) and 99% (outer)
joint confidence region



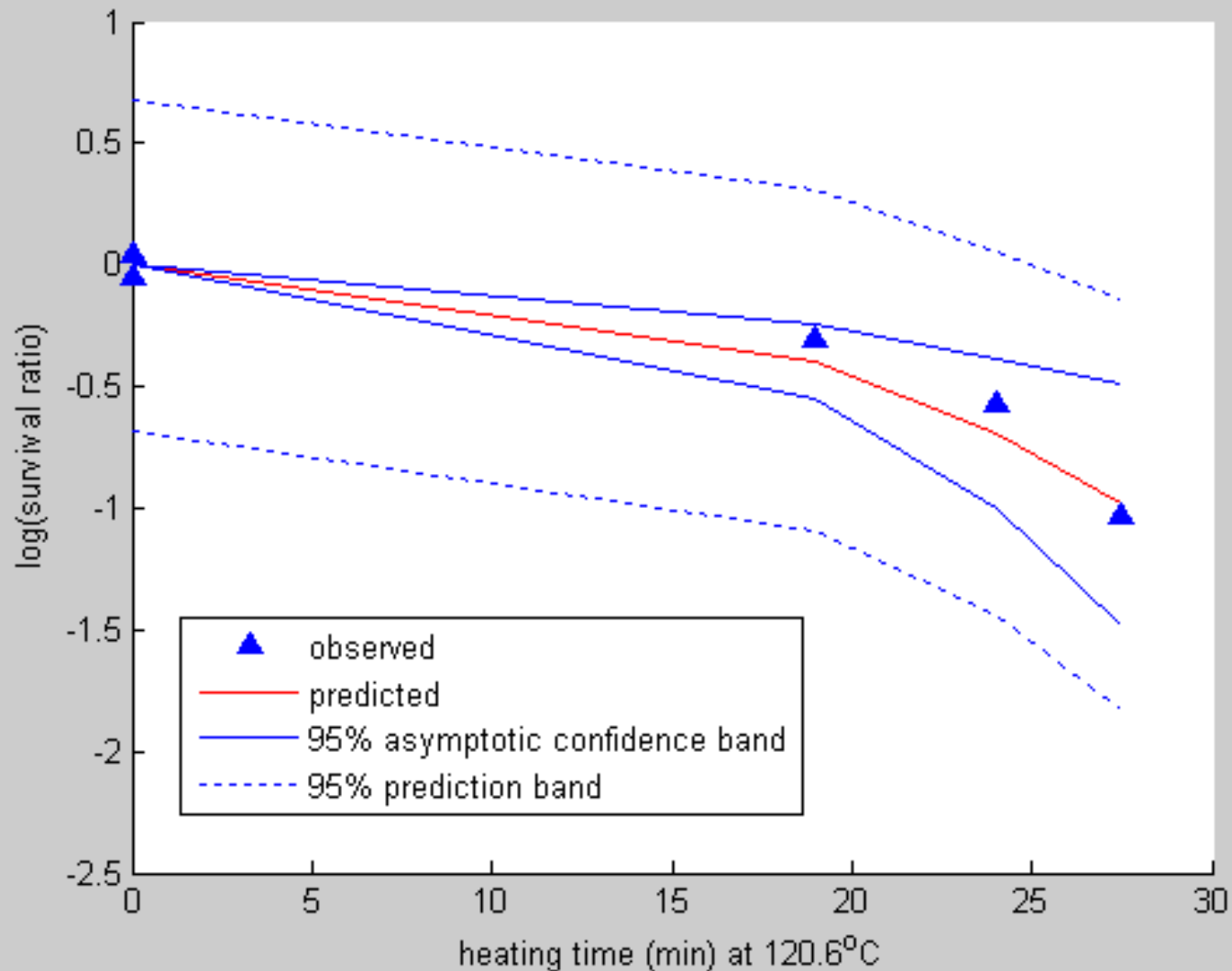
Results @ Retort Temp = 104.4°C



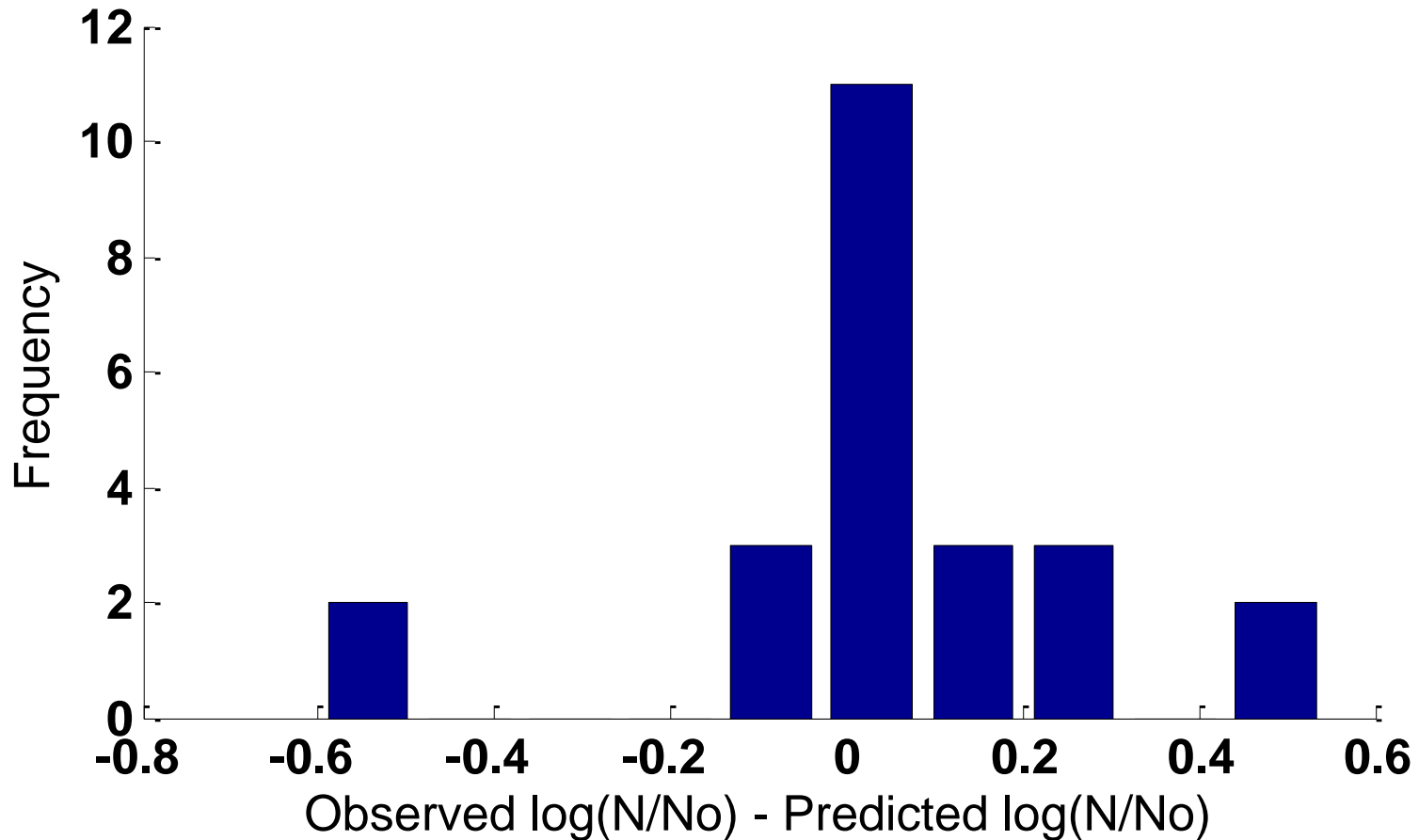
Results @ Retort Temp = 112.8°C



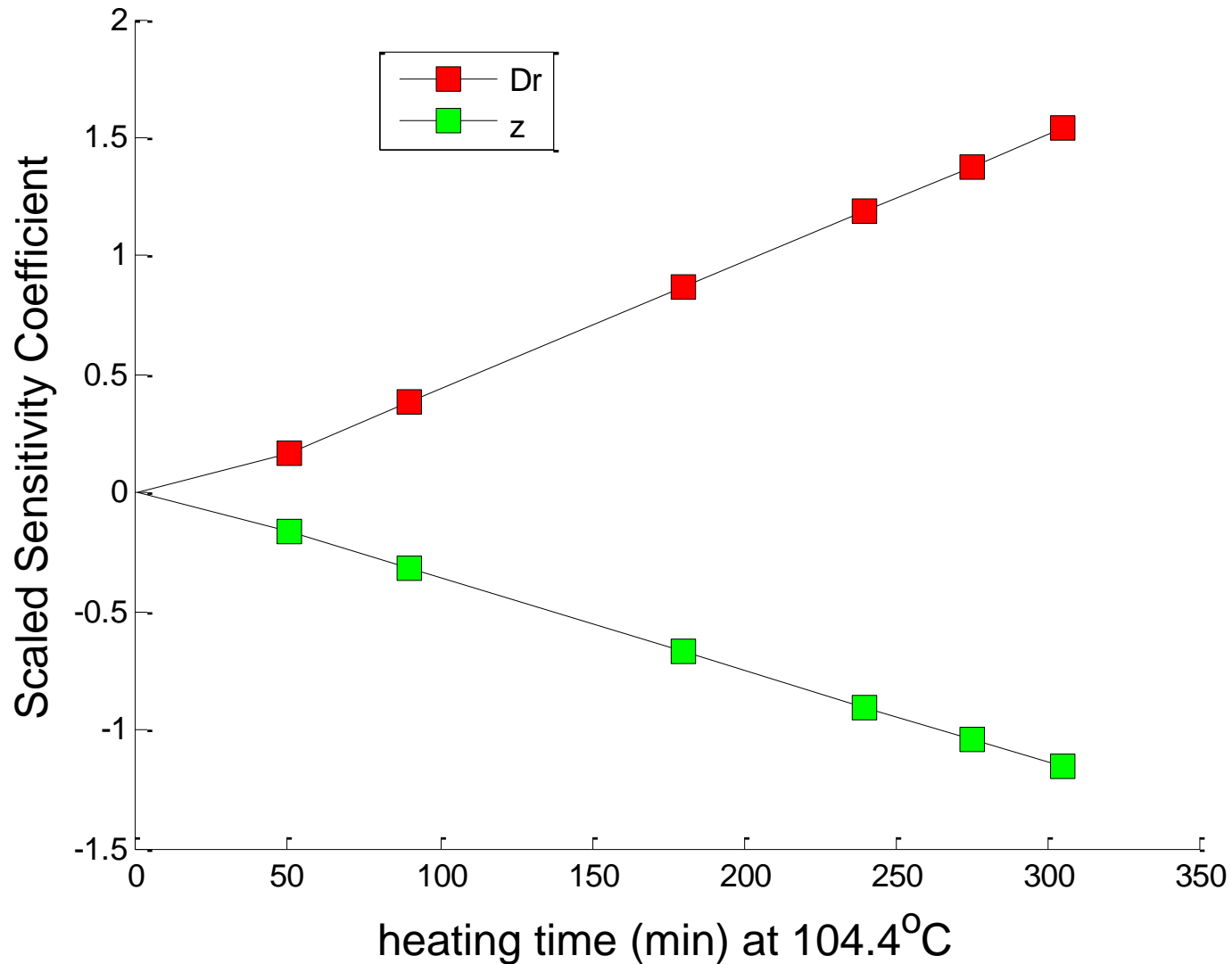
Results @ Retort Temp = 120.6°C



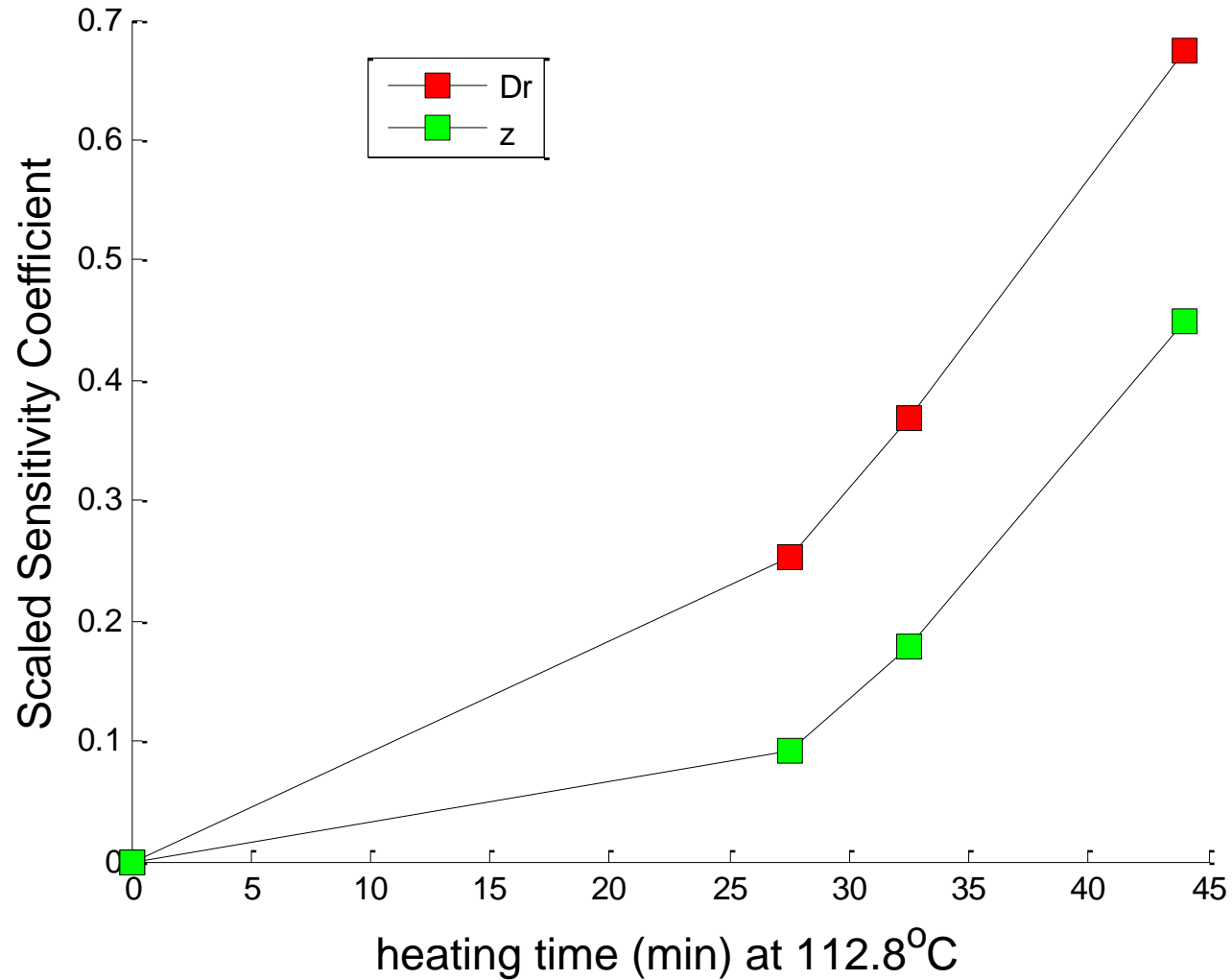
Residual Histogram



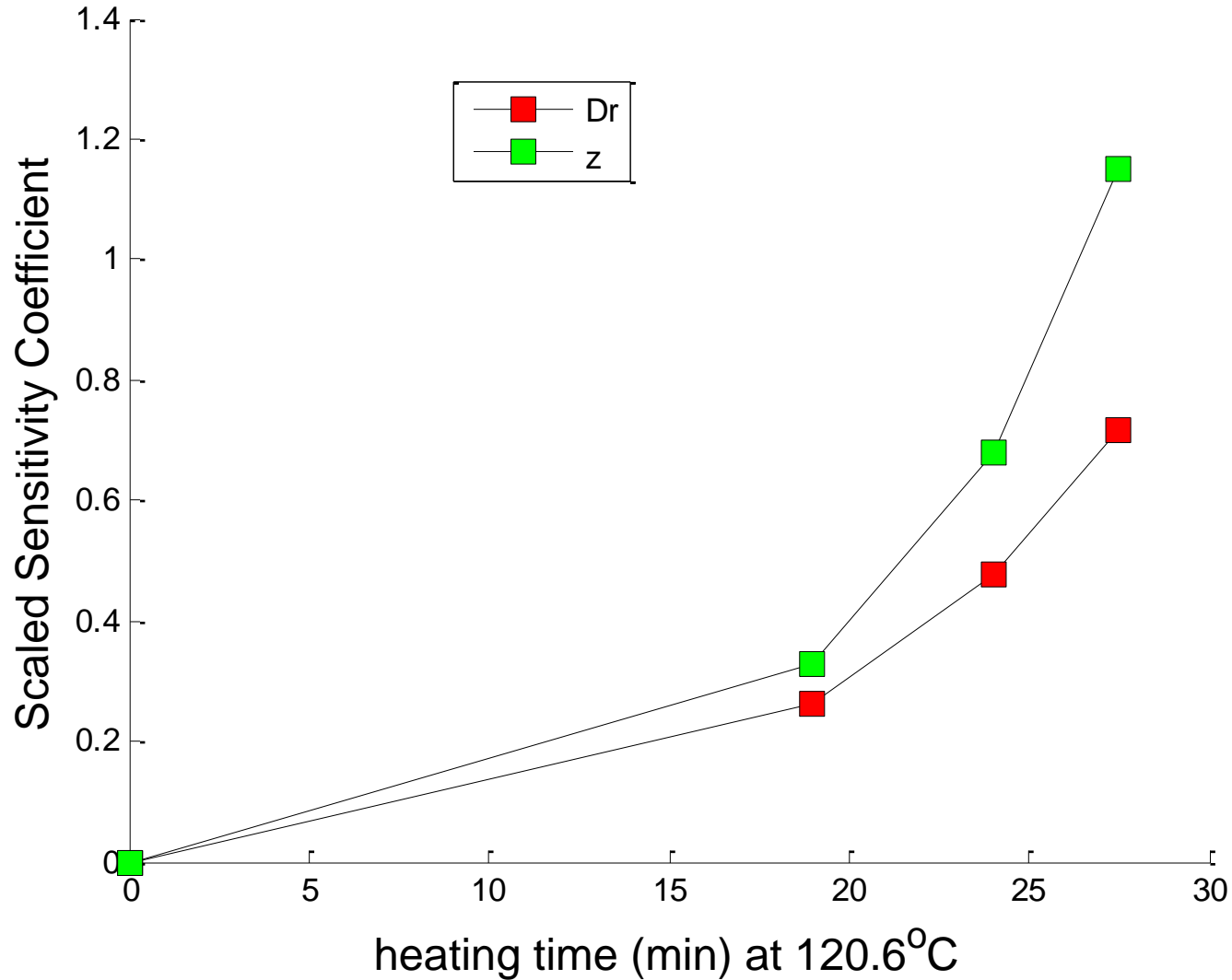
Sensitivity Coefficients at 104.4°C



Sensitivity Coefficients at 112.8°C



Sensitivity Coefficients at 120.6°C



New Information Provided by This Study

- ❑ Microbial kinetic parameters can be estimated simultaneously in conduction-heated foods
- ❑ Confidence bands for the dependent variable were computed
- ❑ Sensitivity coefficients computed
- ❑ Can save experimental \$\$ and effort

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